



Mary Ellen Goldberg BS, LVT, CVT, SRA, CCRA, CVPP

Mary Ellen Goldberg is a graduate of Harcum College and the University of Pennsylvania in 1976. She worked at Virginia Commonwealth University in the Division of Animal Resources and for Research Scientists advising on their choices for anaesthesia and pain management on their protocols. She was a member of VCU's IACUC for 10 years. She has been the instructor of Anesthesia and Pain Management at VetMedTeam, LLC since 2003. She has been the Executive Secretary for the International Veterinary Academy of Pain Management (IVAPM) since 2008. She is a Certified Veterinary Pain Practitioner through IVAPM. Mary Ellen is also a Surgical Research Anesthetist certified through the Academy of Surgical Research. She is on the Exam Chair for APRVT (Academy of Physical Rehabilitation Veterinary Technicians). She is Exam Chair for the Academy of Laboratory Animal Veterinary Technicians and Nurses. Currently, she is a staff member at the Canine Rehabilitation Institute, as a Certified Canine Rehabilitation Veterinary Nurse. (CCRVN). Mary Ellen was chosen as NAVTA's Veterinary Technician of the Year 2017. Mary Ellen has written several books, and contributed to numerous chapters regarding anaesthesia, pain management and rehabilitation. She speaks at national meetings on these topics and gives private CE to organisational groups. She has worked in various aspects of veterinary medicine from small animal and equine to mixed practice, coccidiosis research for a pharmaceutical company, zoo animal medicine and laboratory animal medicine since 1976. Email: mewhitester@gmail.com
DOI: 10.1080/17415349.2018.1529547

A walk on the wild side: a review of physiotherapy for exotics and zoo animals

Mary Ellen Goldberg BS, LVT, CVT, SRA, CCRA, CVPP, VTS-Lab Animal Medicine (Research Anesthesia) VTS-Physical Rehabilitation

6370 Evian Place, Boynton Beach, 33437 Florida, USA

ABSTRACT: When considering physical rehabilitation (physiotherapy) for exotic and zoo animals, it is necessary to recognise painful behaviours in these patients prior to initiation of therapy. Once a creative treatment plan has been formulated, operant conditioning may be utilised in executing the various therapies. This article attempts to document therapies that have been used on various species that do not fall under the category of those normally receiving physiotherapy.

Keywords: physiotherapy; pain; distress; physical rehabilitation modalities; learning

Introduction

Physical Therapy is a protected term in many states in the USA. Only licensed physical therapy professionals may use this term. Success with human patients receiving post-operative physical therapy has galvanised the veterinary community into developing physical rehabilitation techniques that can be implemented for animal patients (Goldberg, 2018). Therefore, the term Veterinary Physical Rehabilitation is commonly utilised in the USA. This terminology may vary based on the country where you practice.

The use of physical rehabilitation for patients other than canine, feline and equine species is individualised. There are not huge amounts of evidence-based literature about exotics and zoo animals. Most literature is either anecdotal or case-based studies. One fact that is known is that the painful animal will be highly resistant to any manipulations that are attempted. With some of these animals, it is dangerous to try anything even when they are not in pain. Thus, recognition of painful patients is necessary (Goldberg, 2017).

Recognition of pain and distress in various species

There are numerous stereotypical responses to stress or pain stimuli in animals, particularly in mammals (Clark, Rager, & Calpin, 1997). Recognition of changes in behaviour and physical appearance in the species under study will allow early identification of an animal experiencing pain or distress. As caregivers, humans may know that an event or situation is no threat, but the animal usually does not function with the same information base as humans.

Personal observations of key signs for various species listed by the author (Table 1). Pain affects every single living creature from pomace flies *Drosophila melanogaster* (Tracey, Wilson, Laurent, & Benzer, 2003) to marine mammals (Haulena & Schmitt, 2018). The nociceptive process has been shown from invertebrates through mammals. The Veterinary Pain Short Course, 2018 (<https://cme.ucsd.edu/vetpain/>) meeting is unique in veterinary medicine for its emphasis

Table 1. Recognition of pain and distress in various species.

Species	Key signs	References
Mice – see Figure 1	Withdrawal, biting response, piloerection, hunched back, sunken eyes and abdomen, dehydration, weight loss	Langford, 2010; Malik & Leach, 2017
Rats – see Figure 2	Vocalisation, struggling, licking/guarding, weight loss, piloerection, hunched position, hypothermia	Carstens & Moberg, 2000; Malik & Leach, 2017
Guinea pigs	Withdrawal, vocalisation, failure to resist restraint, staring coat, unresponsive	Carstens & Moberg, 2000; Gaertner, Hallman, Hankenson, & Batchelder, 2008
Mongolian gerbils	Hunched appearance, weight loss, shock syndrome (fear or surprise may cause convulsions)	Carstens & Moberg, 2000; Gaertner et al., 2008
Syrian (golden) hamsters	Weight loss, hunched appearance, increased aggression or depression, extended sleep periods	Carstens & Moberg, 2000; Gaertner et al., 2008
Rabbits – see Figure 3	Reduced eating and drinking; faces towards back of cage, limited movement, and apparent photosensitivity, tooth-grinding	Johnston, 2005; Carbone & Austin, 2016
Ferrets – see Figure 4	Stiff posture, demented behaviour; lack of grooming, hunched head and neck, and inappetence	Johnston, 2005; Ko & Marini, 2014; Johnson-Delaney, 2017
Birds	Escape reactions, atonic immobility, inappetence and avoidance of use of pain site	Murphy & Ludders, 2001; Lichtenberger & Ko, 2007; Petritz, 2017
Reptiles	Flinching and muscle contractions, weight loss, anorexia	Mosley, 2011; Malik, 2018
Amphibians	Muscular movements, closed eyes, colour changes, rapid respirations, immobility and anorexia	Machin, 1999; Stevens, 2011
Fish	Abnormal swimming behaviour; attempting to jump out of water; rapid opercular movements, clamped fins, pale or darkened colour; hiding. Anorexia is the first sign	Neiffer & Stamper, 2009; Posner, 2009; Sneddon, 2012
Invertebrates	Rapid withdrawal	Mosley & Lewbart, 2014; della Rocca, Di Salvo, Giannettoni, & Goldberg, 2015
Zoo animals – see Figure 5	Behavioural changes, appetite changes, animal isolation from their group or pack, aggression, lameness, unkempt appearance, or lowered head	Whiteside, 2014; Boothe et al., 2016
Non-human primates	Hunched position, failure to groom, refusal of food or water; dejected appearance	Wolfensohn & Honess, 2005; Murphy, Baxter, & Flecknell, 2012
Domesticated species		
Dog – see Figure 6	Whimpers, howls, growls, cowers, crouches, recumbent, reluctant to move; awkward, shuffles	Kata, Roland, & Goldberg, 2015
Cat	Generally silent; may growl or hiss, stiff, hunched in sternal recumbency, limbs tucked under body, reluctant to move limb, carry limb	Kata et al., 2015
Horse	Grunting, nicker, rigid; head lowered, reluctant to move, walk in circles, “up and down” movement, restless, depressed	Kata et al., 2015
Cow	Dull, depressed, inappetence, grunting, grinding of the teeth, and rigid posture	Kata et al., 2015
Sheep	Rigid posture and reluctance to move, tooth-grinding	Kata et al., 2015
Goat	Rigid posture and reluctance to move, tooth-grinding, vocalisation	Kata et al., 2015
Pig	Vocalisation and the lack of normal social behaviour may be helpful indicators of a pig in pain	Kata et al., 2015

upon fundamental (basic) principles and mechanisms of pain as they pertain to the veterinary patient. The 2018 meeting reviewed the evolution of pain behaviour and neuroanatomic substrates. Species discussed went from worms and flies (*drosophila*) to molluscs, cephalopods, reptiles, birds and mammals up to man. The literature clearly shows that while not every animal may have the cognitive abilities of man, they can all certainly respond to an aversive stimulus.

Physical rehabilitation common to canine, feline and equine patients

Physiotherapy is the treatment of injury or illness to decrease pain and restore

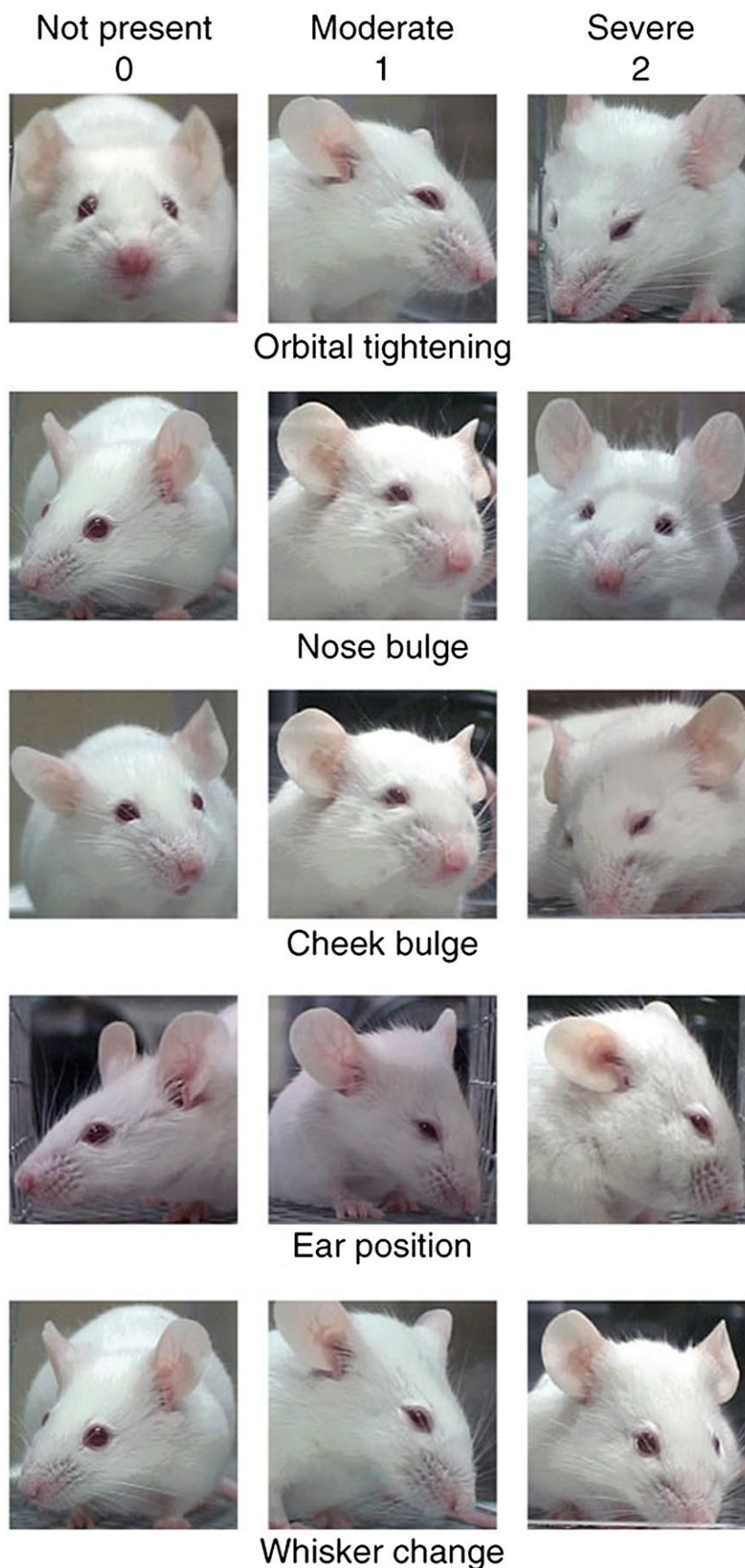
function. Rehabilitation is used to address acute injuries and chronic injuries or diseases that have been affecting a patient for a long time (Goldberg, 2018). Rest alone after injury usually does not relieve the problems caused by inflammation and spasm; for example, a muscle in spasm may not have adequate blood supply to heal. Protective mechanisms in place in the body following injury alter movement of the whole musculoskeletal system and increase strain on other areas. Physical rehabilitation should commence as soon as is possible for the patient and caregiver (Goldberg, 2018).

Details about physiotherapy techniques for the veterinary nurse can be found in Goldberg (2018). The

reader is referred to this textbook for details about each therapy mentioned.

Below are listed the various forms of Physiotherapy used with veterinary patients.

- **Therapeutic exercises** (Coates, Zink, & Van Dyke, 2013) – exercises target proprioception and balance, specific muscle groups, overall pattern of gait and overall strength and endurance. Therapeutic exercise equipment may include physioballs, cavaletti rails, balance blocks and discs, weights, tunnels, rocker boards, wobble boards, treadmills, air mattresses or planks (see Figure 6).
- **Manual techniques** (Millis & Levine, 2014; Saunders, Walker, & Levine,



▲ **Figure 1.** Mouse Grimace Scale progression of pain in mouse face. Langford, 2010.

2014; Sutton & Whitlock, 2014) – techniques used can include joint mobilisations, massage, passive range of motion (PROM), active range of motion (AROM) and stretching. In

the USA, veterinary technicians/nurses are prohibited from performing joint mobilisations; therefore, they must be carried out by the rehabilitation veterinarian or the

rehabilitation-certified physical therapist.

- **Physical modalities** (Niebaum, 2013; Goldberg, 2018) – can be used as tools to manage pain, weak muscles,

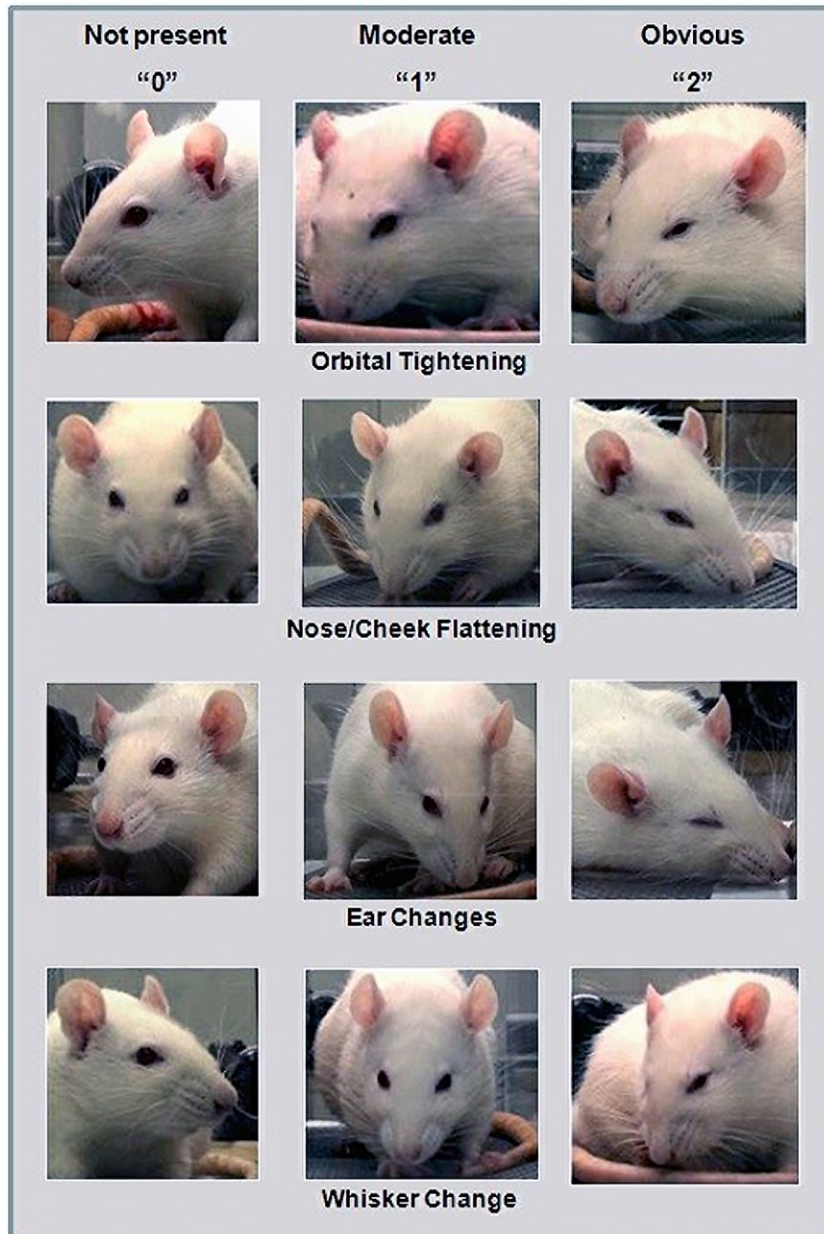


Figure 2. Rat Grimace Scale progression of pain in a rat face. Sotocinal et al., 2011.

inflexibility, limited joint ROM and to aid in tissue healing. Physical modalities include the following:

- superficial thermal agents – hot (thermotherapy) and cold (cryotherapy) (Tomlinson & Goldberg, 2018)
- neuromuscular electrical stimulation (NMES) – usually used to address muscular weakness (Sprague & Goldberg, 2018)
- transcutaneous electrical nerve stimulation (TENS) – used for pain relief (Sprague & Goldberg, 2018)
- therapeutic ultrasound – a deep heating technique used for rehabilitating musculoskeletal conditions (Medina & Davies, 2018)
- low-level laser therapy (LLLT) or therapeutic laser – using (non-surgical) lasers to accelerate wound healing, promote muscle regeneration, treat acute and chronic pain, chronic and acute oedema and neurologic conditions (Shaw & Brown, 2018)
- extracorporeal shockwave therapy (ESWT) – to increase bone, tendon and ligament healing, accelerate wound healing and provide antibacterial properties and pain relief (Stramel & Stramel, 2018)
- pulsed electromagnetic field therapy (PEMF) – to induce biological currents in the tissue. PEMF is approved by the US Food and Drug Administration (FDA) as safe and effective for the treatment of fractures and their sequelae (Rosso, Bonasia, & Marmotti, 2015). The main therapeutic purpose is for enhancement of bone

or tissue healing and pain control (Sprague & Goldberg, 2018).

Learning (Cheng, 2016)

Associative learning is so called because in this type of learning, the animal connects (associates) two types of events. The prevailing view is that one event predicts another event for the animal. In classical conditioning, the animal connects two events in the world, using one event to predict another event. In operant conditioning, the animal learns to connect its own behaviour to consequences. In addition, it needs to learn the circumstances under which a behaviour brings a consequence. That is, a behaviour will “work” only in particular conditions. The appropriate use of operant conditioning can greatly



Figure 3. Rabbit Grimace Scale progression of pain in a rabbit face. Keating, Thomas, Flecknell, & Leach, & Leach, 2012.

	Not present (0)	Moderately present (1)	Obviously present (2)
<p>Orbital tightening</p> <ul style="list-style-type: none"> ▪ The eyelids close (orbital area narrows) ▪ A wrinkle may be visible around the eye 			
<p>Nose bulging</p> <ul style="list-style-type: none"> ▪ The nose is pulled down ▪ The nose rounds off ▪ The nostrils point down ▪ The bridge of the nose bulges 			
<p>Cheek bulging</p> <ul style="list-style-type: none"> ▪ The cheek muscles bulge ▪ The contour of the cheeks become visible ▪ the cheek may be pulled up at the side of the ear 			
<p>Ear changes</p> <ul style="list-style-type: none"> ▪ The ears are pulled back against the body ▪ The ears may form a pointed shape ▪ The ears may fold over 			
<p>Whisker retraction</p> <ul style="list-style-type: none"> ▪ The whiskers are pulled back against the cheek ▪ The whisker follicles converge caudally ▪ The whiskers clump together 			

Figure 4. Ferret Grimace Scale progression of pain in a ferret face. Reijgwart et al., 2017.

facilitate examination and administration of medications when it is used to train zoo animals to reliably exhibit specific behaviours (Crowell-Davis, 2008). In working with zoo animals, positive reinforcement, in which the animal repeats a behaviour because it learns that the behaviour results in a specific, desirable or pleasant consequence, is

most commonly used. The primary operant conditioning technique used to train zoo animals is called shaping. In shaping via positive reinforcement, the animal is reinforced for behaviours that are progressively more and more like the desired end behaviour. Training requires weeks of regular practice and must be maintained through regular

reinforcement of the desired behaviours. Usually, food is the best reinforcer. It must be highly motivating and appropriate to the individual animal and the species (Crowell-Davis, 2008). In the use of physiotherapy for zoo animals, operant conditioning allows them to receive the benefits of the physiotherapy techniques administered, while



▲ **Figure 5.** Elephant depression noticing downcast head and eyes plus lax trunk. Courtesy Stephen Cital.

associating a pleasant reward (mostly food) with the therapy. The discovery of classical conditioning is attributed to the Russian physiologist Ivan Pavlov (1849–1936).

In operant conditioning, often called instrumental conditioning, the animal learns to do things for their consequences. Edward Lee Thorndike (1874–1949) is credited as a pioneer in operant conditioning.

Physical rehabilitation

Creativity is essential for exotic and zoo animal physical rehabilitation. In the case of zoo animals, the zookeeper (person who attends to the daily feed, water, comfort and hygiene of the animal) can be employed in the Home Exercise Program (HEP). Exercises such as weight-shifting, balance blocks and discs, cavaletti rails, physioballs, treadmills and wobble boards

can be taught through use of associative learning: classical conditioning or operant conditioning.

Because creativity is going to play a huge role in physical rehabilitation for exotics and zoo animals, begin by looking at documented studies for what has been used in various species. Most of these studies have been performed in lab animal research.

Mice – experimentally, mice have been used in human research with land treadmills (Al-Jarrah et al, 2007). Running wheels are a good way to increase mouse strength and endurance. Little planks, wobble boards or rocker boards can be made for mice. Swimming experiments have been conducted in mice (Can, 2012).

Cavaletti rails have been made for mice as well as weave cones. Massage

and PROM can be used in mice. Depending upon the size of the laser head, laser therapy has been used in mice experimentally. (Trawitzki, Lilge, de Figueiredo, Macedo, & Issa, 2017).

Rats – experimentally, rats have been used in human research with land treadmills (Selin Cevik, 2018). Little planks, wobble boards or rocker boards can be made for rats. Cavaletti rails have been made for rats as well as weave cones. Massage and PROM can be used in rats. Swimming experiments have been conducted in rats (Alaei, Moloudi, & Sarkaki, 2008). Experimentally, therapeutic laser has been used in rats (de Oliveira, 2018).

Rabbits – experimentally, rabbits have been used in human research with land treadmills (Thu, Kim, & Han, 2017). Equipment used for physical rehabilitation in cats can be utilised on rabbits. Massage and PROM can be used in



▣ **Figure 6.** Dog being worked over cavaletti rails by Mary Ellen Goldberg. Courtesy Mary Ellen Goldberg.

rabbits. Swimming experiments have been conducted in rabbits (Thu et al., 2017). Underwater treadmills have been utilised in rabbits. Experimentally, therapeutic laser has been used in rabbits (Wang et al., 2014).

Ferrets – experimentally, ferrets have been used in human research with land treadmills (Raichlen, Foster, Gerdeman, Seillier, & Giuffrida, 2012). Equipment used for physical rehabilitation in cats can be utilised on ferrets. Massage and PROM can be used in ferrets. Swimming experiments have been conducted in ferrets (Fish & Baudinette, 2008). Experimentally, therapeutic laser has not been used in ferrets; however, guidelines exist in the literature (Mayer & Ness, 2017).

Birds – equipment used for physical rehabilitation in dogs and cats can be utilised

on birds, especially duck and chickens. Cavaletti rails and weave cones have been used in birds. Massage and PROM can be used in birds. Small perching birds can have exercises for their legs and feet with grasping. Laser therapy has been utilised in many bird species like macaws, cockatoos, budgerigars, chicken and ducks (Ness & Mayer, 2017) (Figure 7).

Reptiles – equipment used for physical rehabilitation in dogs and cats can be utilised on turtles or tortoises. Experimentally, treadmills have been used on land and water tortoises (Landberg, Mailhot, & Brainerd, 2003; Pankaew & Milton, 2018). Therapeutic laser has been used on snakes, turtles and lizards (Mayer & Ness, 2017) (Figures 8 and 9).

Aquatic species – Therapeutic laser has been used in aquatic mammals, birds,

invertebrates and fish (Stremme, 2017). In the case of penguins, physical rehabilitation equipment that is used in dogs and cats could be effective. With aquatic mammals such as dolphins (Manire et al., 2018) and manatees (Cray, Dickey, Brewer, & Arheart, 2013) or whales (Smith, Pacini, & Nachtigall, 2018), it is common for them to undergo rehabilitation from injury or stranding with the objective to be released back into the wild. Rehabilitation devices have been made for fish species when they have suffered from swim bladder disease (Anderson 2013; Robson, 2013).

Non-human primates – experimentally, non-human primates have been used in human research with land treadmills (Capogrosso et al., 2016). Equipment used for physical rehabilitation in dogs and cats can be utilised on non-human primates. Massage and PROM can be used in non-human primates.



▣ **Figure 7.** Duck being worked over cavaletti rails. Courtesy Wendy Davies.

No swimming experiments have been conducted in non-human primates. However, swimming pools have been used as environmental enrichment for captive macaques (*Macaca* sp.) (Robins & Waitt, 2011). The swimming behaviour of bonnet macaques has been described (Agoramoorthy, Smallegange, Spruit, & Hsu, 2000). Experimentally, therapeutic laser has not been used in non-human primates; however, guidelines exist in the literature (Boesch, 2012).

Zoo animals – here is where your mind is your limit, so to speak. The author has attempted to provide a brief review of literature for what has been previously attempted.

Physical rehabilitation of a lesser flamingo was successfully accomplished by using swimming and a support sling for developing the ability to regain strength to walk (McEntire & Sanchez, 2017). Progression of therapy consisted of bird being placed in modified sling; two

30-minute swimming sessions added; bird assisted to a standing position every 12 hours; bird standing on its own; use of sling discontinued.

Physical rehabilitation following surgery for angular hind limb deformity of a 2-year-old neutered male pet serval cat is noted (Summa, Eshar, Bichot, Smith, & Moens, 2015). Nutrition was addressed as well as the surgical procedure. A right tibial corrective osteotomy was followed by internal rotation and



Figure 8. Theraband™ use in a Komodo dragon. Courtesy Stephen Cital.



Figure 9. Theraband™ use in a Komodo dragon. Courtesy Stephen Cital.



▣ **Figure 10.** Serval cat in an underwater treadmill. Courtesy Dawn Hickey.

stabilisation with a 2.7mm eight-hole locking compression plate and locking screws. Manipulation of the hind limbs showed no general valgus or varus deformity, but bilateral external rotation of the toes was confirmed and measured with a goniometer. Analgesics, bandaging and mild exercise were prescribed post-operatively. On re-evaluation 4 weeks later, the serval showed marked improvement in posture and gait (Figure 10).

Physical rehabilitation as adjunctive treatment has been performed on a Komodo dragon (Wolfe, et al., 2015). The patient has osteoarthritis that has worsened over the years. It was managed on and off by meloxicam, tramadol and gabapentin until analgesic treatment was no longer effective by itself. A physical therapist was brought in and she chose to use Wolfe Kinetic Technique™, which is a very gentle manual technique that is performed with the intent of changing movement patterns in a way that spreads out the work load of the body when the patient performs functional activities (Wolfe, 2012). In this Komodo dragon, initial treatment was on the thoracic spine, branching out into the scapular stabilisers, lumbar spine and pelvis. Keepers were instructed to perform daily, gentle, manual lateral rocking of the pelvis for 3–5 min. By the tenth treatment, keepers reported that the animal was able to easily negotiate a 12-inch step between

the exhibits, an action which had not been observed for several years.

This case describes the diagnosis and successful treatment of symptoms in a couped leopard (*Neofelis nebulosa*) cub with swimmer-type syndrome (Nájera et al., 2014). The cub presented at 32 days of age unable to adduct both hind limbs and presented hyperextension of both tarsal joints. The treatment focused on gentle flexion and extension movements of the coxo-femoral, femoro-tibial and tibio-tarsal joints. Massaged areas included the main muscle groups of both hind limbs, working the thumb and index fingers from proximal to distal and vice versa. The cub appeared to tolerate the manipulation well and often rested during the procedure. For thermotherapy, warm water-soaked gauze was applied to both hind limbs twice daily for 10-minute periods. To prevent abduction of both hind limbs, a soft gauze bandage was applied as a hobble. At the seventh day of treatment, splints were provided to keep the limbs in normal alignment. All bandages were changed at least once a day and if soiled. After 3 days of splint application, the cub was able to stand and walk with the bandages. After 21 days of treatment, the cub was able to stand, walk and run with no bandages. At 5 months of age the cub was displaying normal locomotive behaviour (Figure 11).



▣ **Figure 11.** Clouded leopard in a hyperbaric oxygen chamber. Courtesy Dawn Hickey.

Laser therapy or photobiomodulation has been described in the literature (Dadone & Harrison, 2017). Operant conditioning is especially beneficial for patients that are too large or dangerous to restrain manually. This therapy has been shown to reduce inflammation, reduce pain and promote healing in laboratory animals and in some domestic species (Draper, 2012; Pryor & Millis, 2015). Treated patients have been described (Dadone & Harrison, 2017) and are:

- Pododermatitis (bumblefoot) in Magellanic penguins (*Spheniscus magellanicus*)
- Chronic pododermatitis in ducks
- Chronic pododermatitis in red-tailed hawk (*Buteo jamaicensis*)
- Wing tip lesions in taptors
- Keel wound in a bald eagle (*Haliaeetus leucocephalus*)
- Pododermatitis in rabbits
- Complications from declaw surgeries in both a caracal (*Caracal caracal*) and a serval (*Leptailurus serval*) healed well when management included laser therapy
- Hygromas in three 2-year-old Amur tigers (*Panthera tigris altaica*) were successfully managed
- Cheek-patch dermatitis in an Asian elephant (*Elaphus maximus*)
- A trunk laceration in an Asian elephant (*Elaphus maximus*)

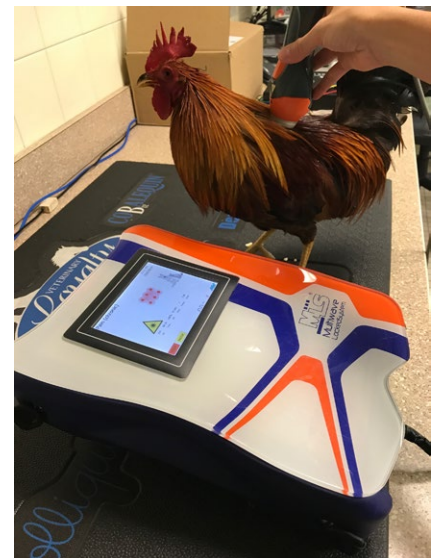


▲ **Figure 12.** Rabbit laser therapy, Courtesy Wendy Davies.

- Wound healing in bearded dragons (*Pogona vitticeps*)
 - Septicaemic ulcerative dermatitis in a soft-shelled turtle (*Pelodiscus sinensis*)
 - Shell lesions in a western pond turtle (*Actinemys marmorata*)
 - Skin lesions in two mata mata turtles (*Chelus fimbriata*)
 - Extensive thermal burns in a savannah monitor (*Varanus exanthematicus*)
 - Lameness in an ostrich (*Struthio camelus*)
 - Acute-onset torticollis in a 2-year-old male reticulated giraffe (*Giraffa camelopardalis reticulata*) showed marked clinical improvement using a combination of medical management, chiropractic adjustments, laser therapy and ROM exercises
 - Nutritional metabolic bone disease and subsequent fractures in a 5-year-old spayed striped skunk (*Mephitis mephitis*)
 - Acute-onset paresis in a Komodo dragon (*Varanus komodoensis*)
 - A spinal cord injury in a bearded dragon (*Pogona vitticeps*) showed clinical improvement with multimodal treatments that included laser therapy
 - A closed distal metacarpal fracture in an orphaned newborn mountain bongo (antelope, *Tragelaphus eurycerus isaaci*)
 - An acute presentation of severe front-leg lameness in an African elephant (*Loxodonta africana*) was managed with multimodal therapies that included laser therapy
 - Four adult or geriatric gorillas (*Gorilla gorilla*) were treated with laser therapy for arthritis pain
- Described is laser therapy for dental disease, musculoskeletal injuries and disease (birds and mammals), such as osteo-arthritis, lumbosacral disease and



▲ **Figure 13.** Rooster laser therapy for congenital feet deformities. Courtesy Dr Carolina Medina.



▲ **Figure 14.** Rooster laser therapy for a deviated spine. Courtesy Dr Carolina Medina.

spondylosis (Dadone & Harrison, 2017) (Figures 12–17).

Conclusion

While massive amounts of published literature are not available, with what has been presented, one can only believe that to use therapeutic exercises, manual therapies and physical modalities on exotics, livestock and zoo patients all depends on the user's imagination. Pain management, acupuncture and physical rehabilitation are evidence-based for companion animals and equine patients. It stands to reason that eventually it will be so for all veterinary patients, no matter the species.



▲ **Figure 15.** A slender-tailed Meerkat receives voluntary laser therapy for arthritis from zookeeper Kirsten Kjorsvig and Janelle Brandt, RVT at the Great Plains Zoo in Sioux Falls, SD. The meerkat enters the box enclosure with inducement from treats. The box enclosure is to hold the meerkat in place for a specific area to receive laser therapy. It is not restrained in this box. Should the meerkat choose to leave the box, it can. This is an example of operant conditioning for a task with a zoo animal.



▲ **Figure 16.** Janelle Brandt, RVT from the Great Plains Zoo in Sioux Falls, SD, performs voluntary laser therapy on a reticulated giraffe for arthritis relief.



Figure 17. Dr Anne Burgdorf and zookeeper Kirsten Kjorsvig from the Great Plains Zoo in Sioux Falls, SD, perform voluntary laser therapy to heal the wound of an eastern black rhino.

Disclosure statement

No potential conflict of interest was reported by the author(s).

References

Agoramoorthy, G., Smallegange, I., Spruit, I., & Hsu, M. J. (2000). Swimming behaviour among bonnet macaques in Tamil Nadu. A preliminary description of a new phenomenon in India. *Folia Primatologica*, 71(3), 152–153. May–Jun

Alaei, H., Moloudi, R., & Sarkaki, A. R. (2008). Effects of treadmill running on mid-term memory and swim speed in the rat with Morris water maze test. *Journal of Bodywork Movement Therapies*, 12(1), 72–75. doi: 10.1016/j.jbmt.2007.05.004. Epub 2007 Jul 20

Al-Jarrah, M., Pothakos, K., Novikova, L., Smirnova, I.V., Kurz, M. J., ... Lau, Y.-S. (2007). Endurance exercise promotes cardiorespiratory rehabilitation without neurorestoration in the chronic mouse model of parkinsonism with severe neurodegeneration. *Neuroscience*, 149(1), 28–37.

Anderson, A. (2013). Retrieved from www.angfaqlid.org.au/aqp/blog/2013/04/24/ada-the-goldfish-beats-swim-bladder-disease/

Boesch, J. M. (2012). Update on primate analgesia. Proceedings of the Annual Conference of the AAZV. Oct 21–26. Oakland, CA.

Boothe, M., Kottwitz, J., Harmon, R., Citino, S. B., Zuba, J. R., & Boothe, D. M. (2016). Results of the megavertebrate analgesia survey: Giraffe and hippopotamus. *Journal of Zoo and Wildlife Medicine*, 47(4), 1049–1056.

Can, A. (2012). The mouse forced swim test. *Journal of Visualized Experiments*, 2012(59), 3638.

Capogrosso, M., Milekovic, T., Borton, D., Wagner, F., Moraud, E. M., Mignardot, J.-B., ... Courtine, G. (2016). A brain–spine interface alleviating gait deficits after spinal cord injury in primates. *Nature*, 539(7628), 284–288.

Carbone, L., & Austin, J. (2016). Pain and laboratory animals: Publication practices for better data reproducibility and better animal welfare. *PLoS One*, 11(5), e0155001. doi: 10.1371/journal.pone.0155001

Carstens, E., & Moberg, G. P. (2000). Recognizing pain and distress in laboratory animals. *ILAR Journal*, 41(2), 62–71.

Cheng, K. (2016). Chapter 3: Basic learning. In *How animals think and feel: An introduction to non-human psychology* (pp. 24–25). Santa Barbara, CA: Greenwood Publications.

Clark, J. D., Rager, D. R., & Calpin, J. P. (1997). Animal well-being II. Stress and distress. *Laboratory Animal Science*, 47(6), 571–579.

Coates, J. (2013). Manual therapy. In M. C Zink & J. B Van Dyke (Eds.), *Canine sports medicine and rehabilitation* (p. 100). Ames, IA: John Wiley & Sons, Inc.

Cray, C., Dickey, M., Brewer, L. B., & Arheart, K. L. (2013). Assessment of serum amyloid A levels in the rehabilitation setting in the Florida manatee (*Trichechus manatus latirostris*). *Journal of Zoo and Wildlife Medicine*, 44(4), 911–917.

Crowell-Davis, S. L. (2008). Use of operant conditioning to facilitate examination of zoo animals. *Compendium Continuing Education for Veterinarians*, 30(4), 218–219.

Dadone, L., & Harrison, T. (2017). Chapter 30: Zoological applications of laser therapy. In R. J. Riegel & J. C. Godbold (Eds.), *Laser therapy in veterinary medicine: Photobiomodulation* (1st ed., pp. 320–333). Ames, IA: John Wiley & Sons.

de Oliveira, H. A. (2018). Protective effects of photobiomodulation against resistance exercise-induced muscle damage and inflammation in rats. *Journal of Sports and Science*, 26, 1–9. doi: 10.1080/02640414.2018.1457419. [Epub ahead of print]

della Rocca, G., Di Salvo, A., Giannettoni, G., & Goldberg, M. E. (2015). Pain and suffering in invertebrates: An insight on cephalopods. *American Journal of Animal and Veterinary Sciences*, 10(2), 77–84.

Draper, W. E. (2012). Low-level laser therapy reduces time to ambulation in dogs after hemilaminectomy: A preliminary study. *Journal of Small Animals and Practice*, 53(8), 465–469.

Fish, F. E., & Baudinette, R. V. (2008). Energetics of swimming by the ferret: Consequences of forelimb paddling. *Comparative Biochemistry and Physiology, Part A, Molecular & Integrative Physiology*, 150(2), 136–143. Epub 2006 Jun 21.

Gaertner, D. J., Hallman, T. M., Hankenson, F. C., & Batchelder, M. A. (2008). Anesthesia and analgesia for laboratory rodents. In R. E. Fish, M. J. Brown, P. J. Danneman, & A. Z. Karas (Eds.), *Anesthesia and analgesia in laboratory animals* (pp. 239–297). San Diego, CA: American College of Laboratory Animal Medicine Series, Academic Press.

Goldberg, M. E. (2017). How to be a pain management advocate for exotic and zoo animals? *The Veterinary Nurse*, 8(7), 389–397.

Goldberg, M. E. (2018). Chapter 1: Introduction to physical rehabilitation for veterinary technicians/nurses. In M. Ellen Goldberg & J. E. Tomlinson (Eds.), *Physical rehabilitation for veterinary technicians and nurses* (1st ed., pp. 1–10). Ames, IA: John Wiley & Sons.

Haulena, M., & Schmitt, T. (2018). Chapter 26: Anesthesia. In F. M. D. Gulland, L. A. Dierauf, & K. L. Whitman (Eds.), *CRC handbook of marine mammal medicine* (pp. 567–606). Boca Raton, FL: CRC Press Taylor & Francis Group

Johnson-Delaney, C. A. (2017). Chapter 24, Analgesia and anaesthesia. In C. Johnson-Delaney (Ed.), *Ferret medicine and surgery* (pp. 377–387). Boca Raton, FL: CRC Press.

Johnston, M. (2005). Clinical approach to analgesia in ferrets and rabbits. *Seminars in avian and exotic pet medicine. Anesthesia and Analgesia*, 14(4), 299–335.

Kata, C. I., Roland, S., & Goldberg, M. E. (2015). Chapter 3: Pain recognition in companion species, horses, and livestock. In M. Ellen Goldberg & N. Shaffran (Eds.), *Pain management for veterinary technicians and nurses* (1st ed., pp. 15–29). Ames, IA: John Wiley & Sons;

Keating, S. C., Thomas, A. A., Flecknell, P. A., & Leach, M. C. (2012). Evaluation of EMLA cream for preventing pain during tattooing of rabbits: Changes in physiological, behavioural and facial expression responses. *PLoS ONE*, 7(9), e44437.

Ko, J. C., & Marini, R. P. (2014). Chapter 12, Anesthesia. In J. G. Fox & R. P. Marini (Eds.), *Biology and diseases of the ferret* (3rd ed., pp. 259–283). Ames, IA: John Wiley & Sons, Ins.

Landberg, T., Mailhot, J. D., & Brainerd, E. L. (2003). Lung ventilation during treadmill locomotion in a terrestrial turtle, *Terrapene carolina*. *Journal of Experimental Biology*, 206(Pt 19), 3391–3404.

- Langford, D. J. (2010). Coding facial expressions of pain in the laboratory mouse. *Nature Methods*, 7(6), 448.
- Lichtenberger, M., & Ko, J. (2007). Anesthesia and analgesia for small mammals and birds. *The Veterinary Clinics of North America. Exotic Animal Practice*, 10(2), 293–315.
- Machin, K. L. (1999). Amphibian pain and analgesia. *Journal of Zoo and Wildlife Medicine: Official Publication of the American Association of Zoo Veterinarians*, 30(1), 2–10.
- Malik, A. (2018). Pain in reptiles: A review for veterinary nurses. *Veterinary Nursing Journal*, 33(7), 201–211.
- Malik, A., & Leach, M. (2017). How do we assess pain in rodents in veterinary practice, what do we know and why it is important? *Veterinary Nursing Journal*, 32(4), 103–108.
- Manire, C. A., Reiber, C. M., Gaspar, C., Rhinehart, H. L., Byrd, L., Sweeney, J., & West, K. L. (2018). Blood chemistry and hematology values in healthy and rehabilitated rough-toothed dolphins (*Steno bredanensis*). *Journal of Wildlife Diseases*, 54(1), 1–13.
- Mayer, J., & Ness, R. D. (2017). Chapter 26: Laser therapy for exotic small mammals. In R. J. Riegel & J. C. Godbold (Eds.), *Laser therapy in veterinary medicine: Photobiomodulation* (1st ed., pp. 287–297). Ames, IA: John Wiley & Sons.
- Mayer, J., & Ness, R. D. (2017). Chapter 28: Laser therapy for reptiles. In R. J. Riegel & J. C. Godbold (Eds.), *Laser therapy in veterinary medicine: Photobiomodulation* (1st ed., pp. 306–312). Ames, IA: John Wiley & Sons.
- McEntire, M. S., & Sanchez, C. (2017). Multimodal drug therapy and physical rehabilitation in the successful treatment of capture myopathy in a lesser flamingo (*Phoeniconaias minor*). *Journal of Avian Medicine and Surgery*, 31(3), 232–238.
- Medina, C., & Davies, W. (2018). Chapter 17: Modalities part 4: Therapeutic ultrasound. In M. Ellen Goldberg & J. E. Tomlinson (Eds.), *Physical rehabilitation for veterinary technicians and nurses* (1st ed., pp. 262–272). Ames, IA: John Wiley & Sons.
- Millis, D. L., & Levine, D. (2014). Chapter 25: Range-of-motion and stretching exercises. In *Canine rehabilitation and physical therapy* (2nd ed., pp. 431–446). Philadelphia, PA: Elsevier.
- Mosley, C. (2011). Pain and nociception in reptiles. *The Veterinary Clinics of North America. Exotic Animal Practice*, 14(1), 45–60.
- Mosley, C. I., & Lewbart, G. A. (2014). Chapter 14, Invertebrates. In G. West, D. Heard, & N. Caulkett (Eds.), *Zoo animal and wildlife immobilization and anesthesia* (2nd ed., p. 204). Ames, IA: John Wiley & Sons, Inc.
- Murphy, K. L., Baxter, M. G., & Flecknell, P. A. (2012). Anesthesia and analgesia in nonhuman primates. In C. R. Abee, K. Mansfield, S. Tardif, & T. Morris (Eds.), *Nonhuman primates in biomedical research volume 1: Biology and management* (pp. 432–433). London, UK: Elsevier/Academic Press, American College of Laboratory Animal Medicine Series.
- Murphy, P. J., & Ludders, J. W. (2001). Avian analgesia. *Veterinary Clinics of North America: Exotic Animal Practice*, 4, 35–45.
- Nájera, F., Brown, J., Kaufman, K., Schwartz, R., Goodrowe, K., Asaithanmakul, W., ... Bush, M. (2014). Swimmer syndrome in a clouded leopard (*Neofelis nebulosa*) cub. *Journal of Zoo and Wildlife Medicine: Official Publication of the American Association of Zoo Veterinarians*, 45(2), 386–388.
- Neiffer, D. L., & Stamper, M. A. (2009). Fish sedation, analgesia, anesthesia, and euthanasia: Considerations, methods, and types of drugs. *ILAR Journal*, 50(4), 343–360.
- Ness, R. D., & Mayer, J. (2017). Chapter 27, Laser therapy for birds. In M. Ellen Goldberg & J. E. Tomlinson (Eds.), *Laser therapy in veterinary medicine: Photobiomodulation* (1st ed., pp. 298–305). Ames, IA: John Wiley & Sons.
- Niebaum, K. (2013). Rehabilitation physical modalities. In M. C. Zink & J. B. Van Dyke (Eds.), *Canine sports medicine and rehabilitation* (pp. 115–128). Ames, IA: John Wiley & Sons, Inc.
- Pankaew, K., & Milton, S. L. (2018). The effects of extended crawling on the physiology and swim performance of loggerhead and green sea turtle hatchlings. *The Journal of Experimental Biology*, 221(1), jeb165225. doi: 10.1242/jeb.165225.
- Petritz, O. A. (2017). No pain, no gain: Updates on analgesia in birds and small mammals. Retrieved from <http://colovma.org/wp-content/uploads/sites/5/2015/11/SA-Petritz.pdf>
- Posner, L. P. (2009). Introduction: Pain and distress in fish: a review of the evidence. *ILAR Journal*, 50(4), 327–328.
- Pryor, B., & Millis, D. L. (2015). Therapeutic laser in veterinary medicine. *The Veterinary Clinics of North America. Small Animal Practice*, 45(1), 45–56.
- Raichlen, D. A., Foster, A. D., Gerdeman, G. L., Seillier, A., & Giuffrida, A. (2012). Wired to run: Exercise-induced endocannabinoid signaling in humans and cursorial mammals with implications for the 'runner's high'. *Journal of Experimental Biology*, 215(8), 1331–1336. doi: 10.1242/jeb.063677.
- Reijgwart, M. L., Schoemaker, N. J., Pascuzzo, R., Leach, M. C., Stodel, M., de Nies, L., ... van Zeeland, Y. R. A. (2017). The composition and initial evaluation of a grimace scale in ferrets after surgical implantation of a telemetry probe. *PLoS ONE*, 12(11), e0187986.
- Robins, J. G., & Waite, C. D. (2011). Improving the welfare of captive macaques (*Macaca* sp.) through the use of water as enrichment. *Journal of Applied Animal Welfare Science: Jaaws*, 14(1), 75–84.
- Robson, S. (2013). Retrieved from www.dailymail.co.uk/news/article-2313439/Disabled-fish-swim-right-way-owner-makes-LIFE-JACKET-stop-sinking-tank.html
- Rosso, F., Bonasia, D. E., & Marmotti, A. (2015). Mechanical stimulation (pulsed electromagnetic fields "PEMF" and extracorporeal shockwave therapy "ESWT") and tendon regeneration: A possible alternative. *Front Aging Neuroscience*, 7, 211.
- Saunders, D. G., Walker, J. R., & Levine, D. (2014). Chapter 26, joint Mobilization. In *Canine rehabilitation and physical therapy* (2nd ed., pp. 447–463). Philadelphia, PA: Elsevier.
- Selin Cevik, O. (2018). Long term treadmill exercise performed to chronic social isolated rats regulate anxiety behavior without improving learning. *Life Science*, 200, 126–133. doi: 10.1016/j.lfs.2018.03.029.
- Shaw, K., & Brown, L. (2018). Chapter 15: Modalities Part 2: Laser therapy. In M. Ellen Goldberg & J. E. Tomlinson (Eds.), *Physical rehabilitation for veterinary technicians and nurses* (1st ed., pp. 231–240). Ames, IA: John Wiley & Sons.
- Smith, A. B., Pacini, A. F., & Nachtigall, P. E. (2018). Modulation rate transfer functions from four species of stranded odontocete (*Stenella longirostris*, *Feresa attenuata*, *Globicephala melas*, and *Mesoplodon densirostris*). *Journal of Comparative Physiology. A, Neuroethology, Sensory, Neural, and Behavioral Physiology*, 204(4), 377–389.
- Sneddon, L. U. (2012). Clinical anesthesia and analgesia in fish. *Journal of Exotic Pet Medicine*, 21(1), 32–43.
- Sotocinal, S. G., Sorge, R. E., Zaloum, A., Tuttle, A. H., Martin, L. J., Wieskopf, J. S., ... Mogil, J. S. (2011). The Rat Grimace Scale: A partially automated method for quantifying pain in the laboratory rat via facial expressions. *Molecular Pain*, 7, 55.
- Sprague, S., & Goldberg, M. E. (2018). Chapter 16: Modalities Part 3: Electrotherapy and electromagnetic therapy. In M. Ellen Goldberg & J. E. Tomlinson (Eds.), *Physical rehabilitation for veterinary technicians and nurses* (1st ed., pp. 241–257). Ames, IA: John Wiley & Sons.
- Stevens, C. W. (2011). Analgesia in amphibians: Preclinical studies and clinical applications. *The Veterinary Clinics of North America. Exotic Animal Practice*, 14(1), 33–44.
- Stramel, D., & Stramel, A. (2018). Chapter 18: Modalities Part 5: Shockwave therapy. In M. Ellen Goldberg & J. E. Tomlinson (Eds.), *Physical rehabilitation for veterinary technicians and nurses* (1st ed., pp. 273–285). Ames, IA: John Wiley & Sons.
- Stremme, D. W. (2017). Chapter 29, Laser therapy for aquatic species. In M. Ellen Goldberg & J. E. Tomlinson (Eds.), *Laser therapy in veterinary medicine: Photobiomodulation* (1st ed., pp. 313–319). Ames, IA: John Wiley & Sons.
- Summa, N. M., Eshar, D., Bichot, S., Smith, D. A., & Moens, N. M. M. (2015). Diagnosis and surgical treatment of a complex angular hind limb deformity in a serval (*Felis serval*). *Journal of Zoo and Wildlife Medicine*, 46(3), 609–612.
- Sutton, A., & Whitlock, D. (2014). Chapter 27, Massage. In *Canine rehabilitation and physical therapy* (2nd ed., pp. 464–483). Philadelphia, PA: Elsevier.
- Thu, V. T., Kim, H. K., & Han, J. (2017). Acute and chronic exercise in animal models. *Advances in Experimental Medicine and Biology*, 999, 55–71. doi: 10.1007/978-981-10-4307-9_4
- Tomlinson, J. E., & Goldberg, M. E. (2018). Chapter 14: Modalities Part 1: Thermotherapy. In M. Ellen Goldberg & J. E. Tomlinson (Eds.), *Physical rehabilitation for veterinary technicians and nurses* (pp. 218–230). Ames, IA: John Wiley & Sons.
- Tracey, W. D., Jr., Wilson, R. I., Laurent, G., & Benzer, S. (2003). *painless*, a *Drosophila* gene essential for nociception. *Cell*, 113, 261–273.
- Trawitzki, B. F., Lilje, L., de Figueiredo, F. A. T., Macedo, A. P., & Issa, J. P. M. (2017). Low-intensity laser therapy efficacy evaluation in mice subjected to acute arthritis condition. *Journal of Photochemical and Photobiology B*, 174, 126–132. doi: 10.1016/j.jphotobiol.2017.07.028. Epub 2017 Jul 26
- Wang, P., Liu, C., Yang, X., Zhou, Y., Wei, X., Ji, Q., ... He, C., (2014). Effects of low-level laser therapy on joint pain, synovitis, anabolic, and catabolic factors in a progressive osteoarthritis rabbit model. *Lasers in Medical Science*, 29(6), 1875–1885.
- Whiteside, D. P. (2014). Chapter 6, Analgesia. In G. West, D. Heard, & N. Caulkett (Eds.), *Zoo animal and wildlife immobilization and anesthesia* (2nd ed., pp. 83–108). Ames, IA: John Wiley & Sons.
- Wolfe, T. C. (2012). Application of the neuroplasticity theory through the use of the Feldenkrais method with a canine with traumatic spinal cord injury: A case study. *Orthopedic and Physical Therapy Practice*, 24, 237–241.
- Wolfe, T. C., Stringer, E., Krauss, S., & Trout, T. (2015). Physical therapy as an adjunctive treatment for severe osteoarthritis in a Komodo dragon (*Varanus komodoensis*). *Journal of Zoo and Wildlife Medicine*, 46(1), 164–166.
- Wolfensohn, S., & Honess, P. (2005). *Handbook of primate husbandry and welfare*. Oxford: Blackwell, 60p.